Tree responses to an invasive sap-feeding insect

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Received: 30 May 2013/Accepted: 10 January 2014/Published online: 25 January 2014 © Springer Science+Business Media Dordrecht 2014

Abstract Although sap-feeding insects are known to negatively affect plant growth and physiology, less is known about sap-feeding insects on woody plants. Adelges tsugae (Annand Hemiptera: Adelgidae), the hemlock woolly adelgid, is an invasive sap-feeding insect in eastern North America that feeds on and kills Tsuga canadensis (L. Carrière), eastern hemlock. Newly hatched adelgid nymphs crawl to young unattacked tissue, settle and immediately enter diapause (aestivation) while attached to hemlock in summer. We assessed the effect of A. tsugae infestation on T. canadensis growth and physiology by analyzing hemlock growth on lateral and terminal branches, water potential, photosynthesis, stomatal conductance, and foliar nitrogen (%N). A. tsugae infestation greatly decreased terminal and lateral growth of eastern hemlock. In addition, A. tsugae presence reduced photosynthesis by 10 % in September and 36 % in October. Adelgid-infested hemlocks also exhibited signs of water stress that included notable reductions in water potential and stomatal

Communicated by Lesley Rigg.

conductance. These responses shed light on possible mechanisms of adelgid-induced mortality.

Keywords $Tsuga canadensis \cdot$ Water potential \cdot Gas exchange $\cdot \%$ Nitrogen \cdot Plant-insect interactions

Introduction

Sap feeders often have long-lasting physiological impacts on their host plant. These physiological changes are driven by both changes in plant nutrients (Masters and Brown 1992; McClure 1980) and the production of secondary chemicals (Haukioja et al. 1990; Karban and Myers 1989). By removing nutrients from the plants' xylem or phloem, sap-feeding insect herbivores decrease plant growth, photosynthesis rates, and reproduction (Candolfi et al. 1993; Meyer 1993). In fact, in some systems sap-feeding insects cause greater damage to woody plants than the defoliators do (Zvereva et al. 2010).

Relatively few studies have investigated the effects of sap-feeding herbivores on woody species (reviewed in Zvereva et al. 2010). Conifers may be especially susceptible to sap feeders because they allocate more storage compounds to foliage as opposed to deciduous trees that store resources in their roots, stems, and other tissues inaccessible to sap feeders (Chapin et al. 1990; Hester et al. 2004; Krause and Raffa 1996). The lack of such stored resources may make conifers more

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Fig. 1 Timing of growth and physiological measurements in relation to the life-histories of *A. tsugae* and *T. canadensis*





likely to succumb to intense sap-feeding events (Fernandes 1990; Furuta and Aloo 1994; Paine 2000). The invasive hemlock woolly adelgid (*Adelges tsugae* Annand Hemiptera: Adelgidae) is a specialist sap-feeding insect currently decimating eastern hemlock (*Tsuga canadensis* (L. Carrière), a foundation species in eastern North American forests. *A. tsugae* feed by inserting their stylet bundle at the base of a needle into the ray parenchyma tissue (Young et al. 1995) and can kill mature hemlocks within 4 years of infestation (McClure 1991). Once *A. tsugae* selects a feeding place, it remains sessile throughout its entire life cycle.

Adelges tsugae has two generations in its invaded range; each generation passes through four larval instars before becoming adults. The April-June progrediens generation lays eggs that hatch in July; the sistens generation that emerges remains on hemlock until the following April when the cycle starts again (McClure 1989). While the progrediens generation feeds continuously throughout their shorter life cycle, the sistens generation enters diapause, a period of arrested growth and metabolic depression, immediately after hatching and settling on young plant tissue with no previous direct exposure to A. tsugae. Summer diapause, also known as aestivation, in A. tsugae is primarily induced by temperature (Salom et al. 2001); Lagalante et al. (2006) suggest that A. tsugae enters diapause to avoid seasonal increases in plant terpenoids.

The aim of this study was to determine how *A*. *tsugae* impacts hemlock anatomy and physiology. Specifically, we assessed hemlock growth during its growing season (April–July) and quantified physiological plant responses as a proxy for host performance (water potential, photosynthesis, stomatal conductance, and foliar nitrogen concentration) once

the foliage had fully matured. We took these measurements in September when the sistens generation is in diapause and in October, immediately after *A. tsugae* resumed feeding (Fig. 1).

Materials and methods

Study site

In April 2011, 1-year-old T. canadensis seedlings $(\sim 0.3 \text{ m in height})$ were purchased from Van Pines (West Olive, MI, USA) and planted in a grid in a randomized complete block design at the Kingston Wildlife Research Station (Kingston, RI, USA). Treatments were randomly assigned within each row of the grid and each row contained every treatment. Trees (n = 15 per treatment) were either experimentally inoculated with A. tsugae or remained uninfested. Insect inoculations were applied in April 2011 and 2012 following a standard protocol (see Butin et al. 2007). Briefly, adelgid-infested hemlock branches collected from naturally infested trees were attached to the trees in the insect treatment while control trees received uninfested branches. Each sapling was enclosed in a cylindrical wire cage (0.3-m diameter and 0.9-m height) covered by a mesh bag (Agribon-15, Johnny's Selected Seeds, Waterville, ME, USA; 90 % light transmission) to prevent cross-contamination. Prior to each physiological sampling (see details below), we counted the number of A. tsugae present/cm of sampled foliage on branches from A. tsugae-infested trees.

Hemlock growth

In April, at the beginning of the plant growing season, two terminal branches and two side branches per tree (n = 15) were marked. For each marked branch, we measured length of new growth starting at bud break (April 28, 2012) and counted the number of buds (starting on June 15) on the newly produced growth (hereafter referred to as "secondary buds"). We measured growth and counted buds every 2 weeks thereafter until July 30, 2012 (when the production of new growth ceased).

Water potential

On September 8 and October 27, 2012 we measured pre-dawn shoot water potential on 12 randomly chosen trees per treatment in the *A. tsugae*-infested and control treatments. Between 4:00 and 5:30 am on each date, we collected one \sim 5 cm cutting from a terminal branch from each tree, wrapped it in a wet paper towel, placed it in a bag, and immediately brought it back to the laboratory in a cooler. Each branch was placed in the pressure chamber of a Scholander pressure bomb (3005 Plant Water Status Console, Soil Moisture Equipment, Santa Barbara, CA, USA) and we recorded the pressure at which xylem appeared visible at the tip of the branch under a magnifying glass.

Gas exchange

We measured gas exchange of new growth (2012) foliage on a terminal branch on each tree used to quantify water potential. Measurements were conducted between 9:00 and 11:00 am on September 9 and October 26, 2012. After each measurement, foliage inside the cuvette was excised and photographed; we quantified total needle area using ImageJ 1.44 software (Abràmoff et al. 2004) to use in gas exchange calculations. To determine gas exchange rates, we used a CIRAS-2 portable photosynthesis system (PP systems, Haverhill, MA, USA) with a 2.5 cm² cuvette and a CIRAS-2 LED light source of 1,500 µmol m⁻² s⁻¹, a CO₂ concentration of 390 ppm, air flow rate at 350 cm³ s⁻¹, and leaf temperature of 25 °C.

Total % nitrogen

We collected new growth needles and stems from 2012 growth for *A. tsugae*-infested (n = 15) and control trees (n = 15) in September and October. These needles and stems were oven dried to constant weight at 70 °C and then finely ground. We

determined total %N through analyzing $\sim 2-3$ mg of each sample by dry combustion with a CHNOS analyzer (vario Micro cube, Elementar Americas, Mt. Laurel, NJ, USA).

Statistical analyses

All statistical analyses were performed using JMP 10.0 with each data point being the mean response variable per tree per sampling date. We used repeatedmeasures ANOVA to analyze the effect of time (repeated factor), treatment, branch type (terminal or side), and all interactions on growth and bud number. We used repeated-measures ANOVA to analyze the main effects of treatment and time, and the treatment*time interaction, on the following variables: water potential, net photosynthesis, stomatal conductance, and %N. We used linear regression to assess the within-treatment correlation between A. tsugae density and water potential, photosynthesis, and stomatal conductance for both time points. We checked all data for normality, homogeneity of variance and sphericity. Water potential data were log transformed in order to meet ANOVA assumptions. For analyses that did not meet the assumptions of sphericity, Greenhouse-Geisser corrected P values are reported.

Results

Hemlock growth

Tree level A. tsugae density averaged 2.16 \pm 0.6 [SE] A. tsugae/cm. A. tsugae infestation had a significant effect on hemlock growth ($F_{1,52} = 7.16$, P = 0.010; Fig. 2a, b) and bud production $(F_{1,52} = 12.34, P = 0.0009;$ Fig. 2c, d). By the end of the growing season, terminal and side branches on control trees were 41 and 57 % longer, respectively, than on A. tsugae-infested trees. There was a significant time*treatment interaction $(F_{6,312} = 136.77, P = 0.0078)$ and time*tissue type interaction ($F_{6,312} = 12.67, P < 0.001$). However, there was no time*treatment*tissue interaction. Branch type had a significant effect on growth and bud production. Terminal branches grew more ($F_{1,52} =$ 16.49, P < 0.001) and produced more secondary buds $(F_{1,52} = 23.78, P < 0.001)$ than side branches. The number of buds increased over time ($F_{3,156} = 2.52$, P < 0.001) and there was a time*tissue interaction



Fig. 2 Average (\pm SE) new growth length and number of new buds in hemlocks infested with *A. tsugae* (*dark gray*) or uninfested (*light gray*) in terminal (**a**, **c**) and side branches (**b**, **d**) from bud break (April) to the end of the growing season (July)

 $(F_{3,156} = 5.22, P = 0.003)$. There were no time*treatment or time*treatment*tissue interactions.

Water potential

Water potential was 45 % higher in control branches than *A. tsugae*-infested branches ($F_{1,20} = 11.36$, P = 0.003; Fig. 3). Water potential changed over time ($F_{1,20} = 7.93$, P = 0.011), but there was no time*treatment interaction ($F_{1,20} = 0.02$, P = 0.900). On both dates, there was a significant negative correlation between *A. tsugae* density and water potential (September: $F_{1,35} = 9.55$, P = 0.006, $R_2 = 0.313$; October: $F_{1,21} = 7.38$, P = 0.010, $R_2 = 0.123$).

Gas exchange

Net photosynthetic rates and stomatal conductance were lower in *A. tsugae*-infested trees ($F_{1,22} = 7.70$



Fig. 3 Average $(\pm SE)$ water potential measurements in *A. tsugae*–infested (*dark gray*) and control trees (*light gray*)

and 14.75, P = 0.011 and <0.001, respectively; Figs 4a, b) compared to control trees. Photosynthesis and stomatal conductance were greater in October ($F_{1,22} = 37.89$ and 111.04, respectively, both P < 0.001). There was a significant time*treatment



Fig. 4 Average (\pm SE) photosynthesis (**a**) and stomatal conductance (**b**) in control (*light gray*) and *A. tsugae*-infested plants (*dark gray*)

interaction for both gas exchange variables ($F_{1,22} =$ 9.43 and 11.35, P = 0.006 and 0.003, respectively). There was a negative correlation between *A. tsugae* density and stomatal conductance for both September ($F_{1,21} = 6.38$, P = 0.020, $R_2 = 0.233$) and October ($F_{1,22} = 7.18$, P = 0.013, $R_2 = 0.238$). There was no relationship between *A. tsugae* density and photosynthesis for September ($F_{1,21} = 1.06$, P = 0.314) or October ($F_{1,22} = 4.02$, P = 0.057).

% Nitrogen

Regardless of insect treatment, %N was higher in needles compared to stems ($F_{1,51} = 200.87$, P < 0.001; Fig. 5a, b). Adelgid-infested foliage had higher %N concentration than control trees ($F_{1,51} = 8.47$, P = 0.0053). Interestingly, %N was greater in infested trees in September but not in October (time*treatment interaction; $F_{1,51} = 12.59$, P < 0.001). When *A. tsugae* were in aestivation (September), %N content was 13 %



Fig. 5 % nitrogen in *A. tsugae*-infested (*dark gray*) and uninfested needles (*light gray*) (**a**) and stems (**b**)

higher in *A. tsugae* -infested needles and 27 % higher in *A. tsugae*-infested stems. However, when *A. tsugae* were actively feeding in October, the %N content differed by less than 1 % for both needles and stems compared to control trees.

Discussion

Our study investigated the effects of an insect's presence on various plant performance traits. We found that *A. tsugae* had significant impacts on hemlock growth. The progrediens generation settles and begins to feed immediately before the start of the hemlock growing season. By the end of the growing season, terminal branches on control trees were 41 % longer and had 56 % more new buds than *A. tsugae*-infested trees. The effect of *A. tsugae* was even more pronounced on side branches. The side branches on uninfested trees grew 56 % more and had 120 % more new buds than *A. tsugae*-infested trees. The significant

treatment*time interactions for both terminal and side branches suggests that A. tsugae has a greater negative impact later in the growing season. While other studies have shown that A. tsugae has a detrimental impact on hemlock growth (McClure 1991; Miller-Pierce et al. 2010), we showed that A. tsugae-infested trees have significantly less lateral growth and buds than uninfested trees. If these trends continue, such difference in branch architecture could have profound effects on small insects with limited active dispersal such as A. tsugae. It is possible that trees with decreased lateral branching will have less branch-to-branch contact, hindering the movement of A. tsugae throughout the tree. In addition, A. tsugae preferentially settle on new tissues so decreased bud production will reduce A. tsugae populations (McClure 1991).

The negative effect of A. tsugae continued even after hemlock growth ceased. Adelgid-infested trees had lower photosynthetic and stomatal conductance rates; this difference was evident in September but was magnified in October. In September, A. tsugae reduced photosynthesis and stomatal conductance by 10 and 29 %, respectively, compared to control trees. In October, however, photosynthesis and stomatal conductance were 36 and 41 % lower in infested trees, respectively. The hemlocks' magnified response to A. tsugae presence in October could be due to the fact that A. tsugae have broken aestivation and are actively feeding by the October time point (Fig. 1). This would be congruent with the idea that sap-feeding insects generally tend to decrease photosynthesis (reviewed in Zvereva et al. 2010) and suggests that this may occur even when a sap-feeding insect is present but inactive. However, it is unclear if the decrease in photosynthetic and stomatal conductance rates is due to the actively feeding A. tsugae, A. tsugae feeding events from prior seasons, or a combination of the two.

The impact of *A. tsugae* may be caused by physical injury or chemical cues associated with the initial stylet insertion. Stylet insertion by other sucking insects has been shown to cause long-lasting anatomical and physiological changes (Ecale and Backus 1995; Ladd and Rawlins 1965; Shackel et al. 2005). *A. tsugae* secretes a salivary sheath when feeding at the base of hemlock needles that Young et al. (1995) suggest may be "toxic" and responsible for the disproportional negative impact on hemlock growth caused by *A. tsugae* feeding. In support of the "toxic" saliva hypothesis, Radville et al. (2011) found that

A. tsugae elicits a local and systemic hypersensitive response in hemlock. Alternatively, the adelgid's impact on photosynthesis and stomatal conductance might be explained by long-lasting changes in nutrient allocation originated when *A. tsugae* began feeding.

Sap-feeding insects often act as resource sinks (Inbar et al. 1995; Kaplan et al. 2011) that can compete with natural plant sinks (i.e., actively growing tissues). This appears to be true when A. tsugae is actively feeding but not when in diapause, as indicated by a 13 % decrease in %N after A. tsugae resumes feeding in October. During diapause, A. tsugae-infested needles and stems have higher %N than control trees; but once A. tsugae begin feeding, %N does not differ between A. tsugae-infested and controls. A. tsugae feeding is known to alter local and systemic foliar nitrogen content (Gómez et al. 2012; Miller-Pierce et al. 2010; Stadler et al. 2005). Nitrogen sink competition between feeding herbivores and new foliage alters leaf nutrient status, resulting in decreased photosynthesis (Larson 1998). Since A. tsugae feed on photosynthate from xylem ray parenchyma cells (Young et al. 1995) that transfer and store nutrients it is likely A. tsugae induce greater sinks when feeding than when in diapause. Although the increase in %N of A. tsugae-infested trees might increase photosynthesis since most leaf N is incorporated into photosynthetic enzymes and pigments (Evans 1989), we instead documented consistent decreases in photosynthesis. A. tsugae feeding increases free amino acids (Gómez et al. 2012), a finding that would explain the increase in total %N found in our study.

High %N (White 1984), low photosynthetic activity, and low stomatal conductance (Dang et al. 1997; Epron and Dreyer 1993; Farquhar and Sharkey 1982) are often linked with water stress. Hemlocks exhibited greater water stress (i.e., more negative water potential) both when A. tsugae were in diapause and feeding. For both time points, this effect was density dependent; water potential was negatively correlated with A. tsugae density. Adelgid-induced water stress may result from the increased formation of false rings in A. tsugae-infested branches (Gonda-King et al. 2012). False rings are bands of abnormal wood within an annual ring that consist of thick-walled xylem cells and which may hinder water transport efficiency (Mitchell 1967). If reductions in water potential are driven by false ring formation, this permanent change in wood anatomy would have a long-lasting impact on water relations and may explain why *A. tsugae* feeding impacts tree physiology.

The *A. tsugae*-hemlock interaction provides an ideal system to study how the presence of sap-feeding insects impact host-plant physiology and performance. Overall, our results suggest that *A. tsugae*-infested trees are water stressed as shown by decreased hemlock growth, water potential, photosynthesis, and stomatal conductance congruent with other hypotheses (Domec et al. 2013). These physiological changes in *A. tsugae*-infested trees may shed light on possible mechanisms behind *A. tsugae*-induced death. We suggest taking long-term measurements on *A. tsugae*-infested trees, from infestation to mortality, to better clarify the mechanism of *A. tsugae*-induced death.

Acknowledgments We would like to thank the RI Audubon Society for access to the Kingston Wildlife Research Station. In addition, the authors thank two anonymous reviewers for insightful comments on the manuscript. LGK was supported by a URI fellowship. This project was funded by an RI-AES Hatch Grant RH-05262 and NIFA 2011-67013-30142 to ELP.

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